

# ***IN SITU* HYBRIDIZATION PROTOCOL**

## ***IN VITRO* TRANSCRIPTION USING DIGOXIGENIN-UTP**

### **DAY 1 OF TRANSCRIPTION:**

1. Add the following to a microtube:

A) 5X TRANSCRIPTION buffer: 4  $\mu$ l

B) RNasin inhibitor : 0.5  $\mu$ l

C) NTP mix: 8  $\mu$ l

Preparation of NTP mix: 3.5  $\mu$ l DIG-UTP

6.5  $\mu$ l UTP (normal)

10.0  $\mu$ l ATP +GTP + CTP each

TOTAL: 40.0  $\mu$ l - vortex and spin.

D) Human Plasmid DNA ( 1 $\mu$ g/ $\mu$ l): 1  $\mu$ l

E) DEPC H<sub>2</sub>O: 4.5  $\mu$ l

F) RNA polymerase: 2  $\mu$ l

TOTAL VOLUME: 20  $\mu$ l

2) Incubate for 2 hours at 37<sup>o</sup>C in waterbath.

3) Add 1  $\mu$ l Rnase-free Dnase to mixture.

4) Incubate in waterbath for 15 minutes at 37<sup>o</sup>C.

5) Add 2.5  $\mu$ l 4 M NACL to mixture.

6) Add 70 ml 100% Ethanol. ( Precool the alcohol to -20<sup>o</sup>C )

7) Precipitate overnight at -20<sup>o</sup>C.

### **Next day:**

1. Spin down at 12000 RPM at 4oC for 10 minutes; discard supernatant.

2. Wash with 70% icecold ethanol 100 $\mu$ l.

3. Spin down 12000 RPM at 4oC for 5-10 minutes; discard supernatant.

4. Dry RNA pellet in speed vacuum for 5 minutes maximum.

5) Resuspend in DEPC H<sub>2</sub>O to 100 ng/ $\mu$ l . Final volume is 20  $\mu$ l.

6) Put for 5 min. at 65 C (to dissolve better ) Store at - 20 C

7) To check the concentration : Run a 2 % gel in 0.5 x TBS .

- a) HAE marker  $0.5 \mu\text{g}/\mu\text{l}$  : use 1  $\mu\text{l}$ .  
DEPC 0.1 % 3  $\mu\text{l}$   
6x Dye 2  $\mu\text{l}$
- b) Unlabelled linearized Plasmid with known conc. 7  $\mu\text{l}$  ( use several dilutions )  
DEPC 0.1 % 3  $\mu\text{l}$   
5 x Dye 2  $\mu\text{l}$
- c) Labelled (linearized) sample : same volumes as b).

This will give you the estimated concentration of your sample.  
You will need 3 ng/section.